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(a-1) Title.

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This should state the results, drawing attention in the text to important details shown in tables and figures.

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This should point out the significance of the results in relation to the reasons for doing the work, and place them in the context of other works. Discussion should not be combined with Results.

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Acknowledgments of people, grants, funds, etc. should be placed in a separate section before the reference list. The names of funding organizations should be written in full.

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References in the text to works by up to three authors should be in full, e.g. (Arakaki & Tokeshi 2011; Yeemin, Sutthacheep & Pettongma 2006). If there are more than three authors, they should always be abbreviated thus: (Susanto *et al.* 2013). When different groups of authors with the same first author and date occur, they should be cited thus: (Darwin, Yucharoen & Samsuvan 1850a; Darwin, Sangmanee & Wongthepwanit 1850b), then subsequently abbreviated to (Darwin *et al.* 1850a, b). The references in the list should be in alphabetical order with the journal name in full. Use the indent function of MS WORD to indent the second and subsequent lines of each entry; do NOT use the space bar to indent - if unsure, do not indent at all. The format for papers, entire books, chapters in books, and PhD theses is as follows.

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 Gomez ED, Aliño PM, Licuanan WRY & Yap HT (1994)
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 (Vol 1) Proc. 3rd ASEAN-Australia Symposium on Living Coastal Resources, Chulalongkorn University, Bangkok.
- Kawai T & Tokeshi M (2007) Testing the facilitation-competition paradigm under the stress-gradient hypothesis: decoupling multiple stress factors. *Proceedings of the Royal Society*, London, B 274, 2503-2508.
- Ota N (2001) Resource utilisation and coexistence in congeneric predatory-scavenging snails, *Japeuthria ferrea* and *J. cingulata*. PhD thesis, Kyushu University, Fukuoka.
- Titlyanov EA, Titlyanova TV & Chapman DJ (2008)
 Dynamics and patterns of algal colonization on mechanically damaged and dead colonies of the coral *Porites lutea. Botanica Marina* 51, 285-296.
- Tokeshi M (1999) Species Coexistence: Ecological and Evolutionary Perspectives. Blackwell Science, Oxford.
- Yap HT, Alino PM & Gomez ED (1992) Trends in growth and mortality of three coral species (Anthozoa: Scleractinia), including effects of transplantation.
 Marine Ecology Progress Series 83, 91-101.

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1	Regular paper
2	
3	Distribution and settlement of <i>Ruditapes philippinarum</i> in the Suo-Nada Sea, Japan
4	
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6	
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17	Running headline: Manila clam larval distribution and settlement
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23	

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(<mark>Abstract should be on page 2</mark>)

25 Abstract

26Understanding the causes of spatiotemporal variations in the scale of larval settlement is 27important for population dynamics studies in bivalves. This study investigated the seasonal 28abundance of the asari or Manila clam (Ruditapes philippinarum) larvae over a 4-year period (2004–2007) and their settlers over 3 years (2005–2007) in the Suo-Nada Sea, Japan. 2930 Seasonal differences in larval transport were examined by numerical simulation using 3D 31ocean-modelling. During the 2004–2007 spawning seasons, larval numbers peaked two or 32three times in June/July, August/September, and October/November. Settler occurrence was 33uncoupled with larval occurrence; settler density was >10 times higher in October/November than in other months. Numerical simulation suggested that the extent of larval transport 3435differed seasonally; larval transport via loss from the Suo-Nada Sea was estimated to be 20% 36 in June/July, whereas it was almost 0% in November. However, this could not explain the 37seasonal difference (>10-fold) in settler density. In addition, the average density of larger 38larvae (>180 μm) during 2004–2007 was higher in June/July than in October/November, as a 39result of spawning and larval loss (via transport and mortality), suggesting that larval supply 40 alone could not explain the seasonal differences in settler density. These results suggest that 41the seasonal differences in settler density were affected more by variation in mortality during 42the settlement and/or early post-settlement stages, which may depend on environmental 43conditions at the settling site, rather than by larval supply.

44

45 Keywords: clam, larval supply, larval transport, settlement

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47 Introduction

48

49	For many marine bivalves including the asari or Manila clam Ruditapes philippinarum
50	(Adams & Reeve 1850), spatio-temporal differences in larval supply constitute a potential
51	cause of variations in the scale of larval settlement (Young et al. 1998; Shanks & Brink
52	2005). Understanding the causes of variations in larval supply and settlement, along with
53	post-settlement processes, is important for studies on the population dynamics of bivalves
54	(Hunt & Scheibling 1997; Pineda, Hare & Sponaugle 2007).
55	R. philippinarum was originally distributed in the temperate to subarctic regions
56	along the east coast of the Pacific Ocean, but is currently found in many other areas
57	worldwide because of its introduction for aquaculture (Goulletquer 1997). Population studies
58	on R. philippinarum have attracted attention because of its commercial and ecological
59	importance, especially after a population decline in Japan (Miyawaki & Sekiguchi 1999; Ishii
60	et al. 2001; Toba et al.2007; Tamaki et al. 2008; Tezuka et al. 2012). Spawning and larval
61	occurrence of the asari clam has been observed during spring to autumn at water
62	temperatures above 14 °C (Miyawaki & Sekiguchi 1999; Ishii et al. 2001; Matsumura et al.
63	2001; Drummond, Mulcahy & Culloty 2006). Dense larval settlement has often been
64	recorded in autumn in temperate regions of Japan (Miyawaki & Sekiguchi 1999; Ishii et al.
65	2001; Toba et al. 2007). However, the causes of seasonal variation in settler density and the
66	decoupling between larval occurrence and settlement are as yet unclear.
67	The Suo-Nada Sea was renowned as one of the major R. philippinarum fishery
68	ground in Japan until the population drastically declined in the late 1980s (see Tezuka et al.
69	2012, 2014). Surveys conducted in the 1970s and early 1980s, before the population decline,
70	showed that larval abundance peaked in spring and autumn, and that settlers were observed
71	mainly in autumn (Inoue 1980; Fujimoto et al. 1985). No larval surveys have been carried

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72	out after the population decline; however, seasonal observations of settlement on the Nakatsu
73	tidal flats in the Suo-Nada Sea have been reported recently by Tezuka et al. (2012), and
74	settlers have been observed in autumn.
75	To ascertain the causes of seasonal differences in settler density observed on the
76	Nakatsu tidal flats during 2005–2007 by Tezuka et al. (2012), we investigated the abundance
77	and distribution of R. philippinarum larvae between April and November over the period
78	2004–2007. In addition, seasonal differences in larval transport were examined by numerical
79	simulation. Data for settler numbers were obtained from the Tezuka et al. (2012) study and
80	reanalyzed to determine the periods of larval settlement using shell-length distribution. The
81	study will increase our understanding of the causes of seasonal differences in settler density
82	and larval numbers after the clam population decline in the Suo-Nada Sea.
83	
84	Materials and Methods
85	
86	Sampling of larvae and settlers

87 Sampling of R. philippinarum larvae was conducted in the Suo-Nada Sea between April and November over the period 2004–2007 (Fig. 1). Larvae were collected by pumping 200 L of 88 89 seawater from a 5-m depth and screening onto a 50-µm mesh net. The sampling depth was 90 changed to an intermediate level in cases where the sampling stations were shallower than 10 91m. In the periods 19-22 June 2007, 20-23 August 2007, 15-19 October 2007, and 16-19 92November 2007, multilayer sampling from 2-, 5- and 10-m depths was conducted to ascertain 93the vertical distribution of clam larvae. Samples were frozen until needed for identification, 94enumeration and shell length measurement. After thawing, larvae were identified using 95fluorescent antibodies and counted under a fluorescence microscope (Matsumura et al. 2001;

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96	Kasuya, Hamaguchi & Furukawa 2004). Shell length was measured using an eyepiece to the
97	nearest 10 μ m, and up to 100 individuals were measured per sample.
98	R. philippinarum settlers were collected at two stations on the Nakatsu tidal flats
99	during the period 2005–2007 (Fig. 1; see Tezuka et al. 2012) by taking three, 1-cm deep
100	surface-sediment samples with a core sampler (40 mm in diameter). Samples were preserved
101	by freezing until settlers were counted and shell lengths measured to the nearest 0.1 mm.
102	
103	Numerical simulation of larval transport
104	A 3D numerical ocean model was used for simulation of <i>R. philippinarum</i> larval transport in
105	the Suo-Nada Sea. Particle-tracking experiments were conducted using the same flow-field
106	reported by Chang et al. (2009). A total of 1000 modelled larval particles were set in each of
107	nine areas along the Suo-Nada coast (Fig. 2) and tracked by the Euler-Lagrange method for
108	14 days (100 particles per 10-model grids per area). The simulation was conducted over two
109	seasonal periods: June/July (from June 30 to July 17) and November (from November 7 to
110	November 27).

111 The position of the particle $X_{n+1}(x^{n+1}, y^{n+1})$ at time n+1, which was $X_n(x^n, y^n)$ at time 112 *n*, can be calculated by the following equation:

113
$$X_{n+1} = X_n + V\Delta t + (\partial V/\partial t + V\nabla V)\Delta t^2/2 + R$$

114 where *V* denotes the horizontal velocity vector of residual flow; Δt , the time step; ∇ , the 115 horizontal gradient. *R* is the dispersion due to turbulence and is given by the following 116 equation:

117
$$R = \gamma (2\Delta t D_h)^{1/2}$$

118 where γ is the normal random number whose average is zero and whose standard deviation is 119 1.0. D_h is the horizontal dispersion coefficient that depends on Smagorinsky diffusivity 120 (Mellor 2004), which varies with horizontal shear stress.

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121	Vertical migration of larval particles was hypothesized as larvae were located at a
122	3-m depth from day 0 to day 11 but, then moved to 1 m above the ocean floor between day
123	12 and day 14. This assumption for the vertical migration of larval particles is based on
124	observational studies on the vertical distribution of larvae, including this study (see Results,
125	Fig. 6), which showed that smaller larvae (D- and umbo-shaped) were found at \sim 3 m,
126	whereas larger larvae (settling larvae) found in the bottom layer (Suzuki et al. 2002; Ishii,
127	Sekiguchi & Jinnai 2005; Kuroda 2005; Toba et al. 2012; Bidegain et al. 2013). The larval
128	stage was assumed to last 2 weeks (14 days) in this study, although it can vary with
129	temperature and food availability (Helm & Bourne 2004).
130	The larval retention rates, i.e., the ratio of larval particles remaining within the
131	Suo-Nada Sea, after the 14-day simulation were calculated as follows:
132	Retention rate after 14 days = $100 \times P_a / P_0$
133	where P_a is the number of particles remaining within the Suo-Nada Sea after the 14-day
134	simulation, and P_0 is the number of particles within the Suo-Nada Sea on Day 0. The
135	boundary of the Suo-Nada Sea was set on a line through two points (131.7°E, 33.7°N) and
136	(132.0°E, 34.0°N) (see Fig. 1), and particles located to the western side of the boundary line
137	were treated as inside the Suo-Nada Sea. Retention rates were calculated for each of the nine
138	areas where particles were released for two seasonal periods.
139	
140	Results
141	
142	Seasonal dynamics of larval distribution and settlement

143 Distribution of *R. philippinarum* larvae from April to November 2004–2007 is shown in Fig.

- 144 3. Seasonal changes in planktonic larval abundance are shown in Fig. 4. *R. philippinarum*
- 145 larvae were observed sporadically from April to November, with peak numbers being

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146	recorded two or three times in a year, i.e. June/July, August/September and
147	October/November (Figs. 3 and 4). Larvae were distributed more widely in the Suo-Nada Sea
148	during June/July (e.g. 13–16 June 2005, 19–2 June 2006 and 19–22 June 2007) than in
149	October/November (e.g. 20-23 November 2005, 30 October-2 November 2006 and 15-19
150	October 2007) (Fig. 3). The maximum recorded planktonic larval density was c. 2000
151	individuals m ⁻³ , observed in November 2006 and in June 2007 (Fig. 4).
152	The size-frequency distribution of <i>R. philippinarum</i> settlers on the Nakatsu tidal
153	flats between May–December 2005–2007 is shown in Fig. 5. New settlers (< 0.5 mm in shell
154	length) were found almost exclusively in October/November for the 3-year period (Fig. 5).
155	The density of new settlers (< 0.5 mm in shell length) differed by more than 10-fold between
156	summer and autumn: 200-1000 individuals m ⁻² in June–August, and 4000-13,000 individuals
157	m ⁻² in October/November.
158	Vertical distribution of <i>R. philippinarum</i> larvae, inferred from the multilayer
159	sampling trials, are shown in Fig. 6. During 19–22 June 2007, smaller larvae (120–180 μ m in
160	shell length) were more abundant at the 2- and 5-m depths than at 10-m, whereas larger
161	larvae (>180 μ m) were more abundant at 10 m. During the other time-periods, larvae were
162	less abundant than during 19–22 June 2007, being found almost exclusively at 2 m.
163	

164 Numerical simulation of larval transport

Results of the numerical simulation of larval particle-tracking are shown in Fig. 7. In the June/July period, released larval particles were carried extensively by two major currents that developed in the Suo-Nada Sea; a circular current in the central region and a northward current along the western coast. In the November simulation, the currents had weakened and the larval particles were retained within the Suo-Nada Sea. The retention rates for particles within the Suo-Nada Sea after the 14-day simulations for June/July and November are shown

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171	in Table 1. In the June/July period, retention rates of larval particles varied by area where the
172	particles were released, with rates ranging from 32.2% (Area 4) to 100% (Areas 5 and 6). On
173	average, 79.5% of the particles were retained within the Suo-nada Sea, while the rest were
174	lost by being transported from the Suo-Nada Sea in the June/July period. For the November
175	period, retention rates were higher than in the June/July period at >99%.
176	The densities of planktonic larvae of different sizes are compared between June/July
177	and October/November for combined data (2004–2007) (Fig. 8). The average larval density
178	of all combined size classes was 79.2 individuals m^{-3} in June/July and 36.3 individuals m^{-3} in
179	October/November, i.e., approximately twice as higher in the former. Of these, larger larvae
180	(>180 μm) constituted 8.0% in June/July (6.3 individuals $m^{\text{-3}}$) and 10.2% in
181	October/November (3.7 individuals m ⁻³).
182	
104	
183	Discussion
	Discussion
183	Discussion This study reported the seasonal dynamics of <i>R. philippinarum</i> larval distribution in the
183 184	
183 184 185	This study reported the seasonal dynamics of <i>R. philippinarum</i> larval distribution in the
183 184 185 186	This study reported the seasonal dynamics of <i>R. philippinarum</i> larval distribution in the Suo-Nada Sea and the number of settlers on the Nakatsu tidal flats. Larval occurrence and
183 184 185 186 187	This study reported the seasonal dynamics of <i>R. philippinarum</i> larval distribution in the Suo-Nada Sea and the number of settlers on the Nakatsu tidal flats. Larval occurrence and settlement events were not necessarily coupled and, although larvae were recorded from
183 184 185 186 187 188	This study reported the seasonal dynamics of <i>R. philippinarum</i> larval distribution in the Suo-Nada Sea and the number of settlers on the Nakatsu tidal flats. Larval occurrence and settlement events were not necessarily coupled and, although larvae were recorded from April to November, settlement occurred almost exclusively in October/November. Numerical
183 184 185 186 187 188 189	This study reported the seasonal dynamics of <i>R. philippinarum</i> larval distribution in the Suo-Nada Sea and the number of settlers on the Nakatsu tidal flats. Larval occurrence and settlement events were not necessarily coupled and, although larvae were recorded from April to November, settlement occurred almost exclusively in October/November. Numerical simulation suggested that larval loss via current transport was higher in June/July than in
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similar to that reported in previous studies within the Suo-Nada Sea and other regions. Inoue(1980) reported that planktonic larval density peaked in May and October/November in 1976

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196 on the northern coast of the Suo-Nada Sea (maximum larval density was 14,000 individuals m^{-3} in October 1976), and that new settlers were found in abundance (26,700 individuals m^{-2} 197 198in November 1976) in October/November, but very few, new settlers were recorded in other 199 months. Fujimoto et al. (1985) observed that larval density peaked in April, June/July, 200September and November during the period 1983–1984 on the southwest coast of the Suo-Nada Sea, with a maximum density of 4550 individuals m⁻³ in November 1983 and 6140 201individuals m⁻³ in April 1984 (larval density was calculated as the mean from eight sampling 202203 stations). An increase in new settlers was observed in October/November 1983 (maximum of 204350.000 individuals m⁻² in November 1983) and in November/December 1984 (max. 20.000 individuals m⁻²). Although the maximum larval density observed in the current study was 205206lower than those recorded in the 1970s and 1980s, the seasonal pattern was consistent among 207these studies. The difference in maximum larval density could be a result of differences in 208the spawning biomass between the 2000s and the 1970–1980s; clam production in the 209 Suo-Nada Sea was 14,800 metric tonnes in 1976, 19,500-31,700 metric tonnes in 1983/1984, 210and 94-780 metric tonnes during 2005-2007 (Ministry of Agriculture, Forestry and Fisheries, 211Japan, 2012). Dense larval settlement in autumn has often been observed in other temperate 212regions of Japan (Miyawaki & Sekiguchi 1999; Ishii et al. 2001; Toba et al. 2007). 213The results of numerical simulation suggest that larval retention in the Suo-Nada Sea 214should be higher, i.e., lower larval loss rates, in October/November than June/July, owing to 215seasonal differences in the strength of two major currents in the Suo-Nada Sea. As 216 stratification developed in June/July, a circular current developed in the central region and a 217northern current developed along the southwest coast of the Suo-Nada Sea. However, as 218stratification declined in October/November, the currents weakened. The simulation 219suggested that larval loss via transport from the Suo-Nada Sea was 20% in June/July and 0%

in October/November. However, this disparity could not explain the >10-fold difference in

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settler density between October/November and June/August. Moreover, on average during
the period 2004–2007, the observed density of >180 µm larvae was higher in June/July than
in October/November. This suggests that, even if he larval loss via transport may be higher in
June/July, larval supply, as a result of spawning and larval loss via transport and mortality,
was not the limiting factor in June/July under the prevailing conditions.

226Factors influencing settler density, other than larval supply, need to be considered, 227and although not all factors were investigated in this study, several hypotheses may be 228proposed. First, mortality during the critical period from settling to early post-settlement 229affects settler density (Hunt & Scheibling 1997), and this mortality may be lower in 230October/November than in other months. Predation and/or other factors, such as high 231temperature in summer, may cause higher mortality in settling larvae or early settlers during 232months other than October/November. Second, food availability for larvae may differ 233seasonally, resulting in differences in mortality during settlement (Laing 1995; Tezuka et al. 2342009). Third, the rate of larval settlement may vary due to seasonal differences in physical 235conditions at settling sites, e.g., variation in sediment grain size (Tezuka et al. 2013). If a 236suitable substrate is not found, larvae could delay settlement (Coon, Fitt & Bonar 1990) and, 237as a result, become more prone to predation by benthic filter-feeders while drifting near the 238bottom (Pineda et al. 2010). This study concentrated solely on larval settlement on the tidal 239flats and other possible sites, e.g., subtidal areas, were not investigated. As such, settlement 240failure on the tidal flats in seasons other than October/November needs to be investigated 241further.

242

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339	

Table 1 Retention rate (%) of particles within the Suo-Nada Sea after a 14-day simulation. Retention rates were calculated for each of the nine area where particles were released over two seasonal periods, June/July and November.

Area	1	2	3	4	5	6	7	8	9	Ave.
June/July	85.4	93.7	40.4	32.2	100	100	75.6	89.9	98.7	79.5
Nov	99.7	100	100	100	100	100	100	100	99.4	99.9

(Each Table should be on a separate sheet.)

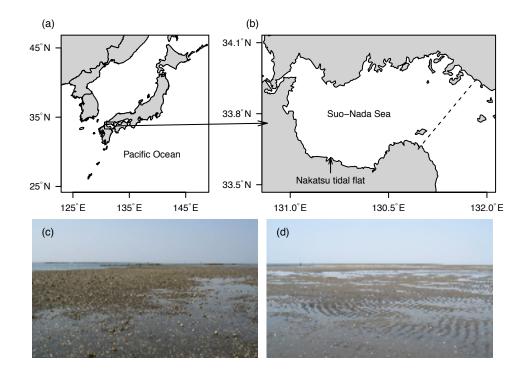
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340 Figure legends

341

- Fig. 1 (a)-(b), Map of the Suo-Nada Sea, Japan. Dashed line in (b) is the boundary used to
- 343 calculate the larval retention rate (see text). (c)-(d), Nakatsu tidal flats where larval settlement
- was recorded.
- 345 Fig. 2 Modelled larval particles were set in nine areas along the Suo-Nada Sea coast for
- 346 numerical simulation of *Ruditapes philippinarum* larval transport. A total of 1000 modeled
- 347 larval particles were set on 10-model grids in each area. The model grids in each area were
- 348 indicated as "+" symbols with identical color.

(other figures not shown in this example)





(Note: Figure originals should NOT have figure number.)

